

Express test for the simultaneous diagnosis of encountering PAMG-1 and PAMG - 2 using technique of immune chromatography using nanozolota and IPMS-ELISA using isotopes Ei^{3+} and Sm^{3+}

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Summary: the simultaneous determination of PAMG-1 and PAMG — 2 in blood serum by enzyme immunoassay (and PMS-ELISA) based on inductively coupled plasma mass spectrometry (IPMS-ELISA) and immunochromatographic analysis was studied. The PAMG-1 and PAMG-2 measurement ranges were 1-100ng/ml, respectively, with detection limits I (3 SD above the mean of the zero calibrator), respectively. Compare and PMS-OLD ECHO for PAMG-1 and PAMG — 2 have correlation coefficients (r^2) of 0.97 and 0.95. Thus, it is shown that both methods allow to measure two proteins with comparable accuracy.

Key words: ELISA, PAMG-1, PAMG-2, Ei^{3+} , Sm^{3+} , immunochromatography

Simultaneous multianalysis of several analytes is important for solving a wide range of biological and medical problems, including for screening diagnosis of cancer pathology. The use of conjugates of gold nanoparticles, quantum dots and other achievements of nanotechnology in recent years can effectively solve the problem of multianalysis. For these goals, developing hybrid methods, including ELISA, radioanalyt, differential counting multimeter. Such analysis, for example, is the IPMS-ELISA method [1,2]

Materials and methods

PAMG-1 and PAMG-2, and monoclonal mouse antibodies against them were obtained from Nano-lab LLC, Russia. We used deionized water with a resistance of 18 MW for all experiments. N^{\prime} — [p-isothiocyanatobenzyl] — diethylene-triamine-H1,H2, N3, N3-tetraacetate europium and N-[p— isothiocyanato-benzyl]-monoethylene glycol — triamine-H1,H2, N3, N3-samarium tetraacetate are obtained from Tianjin radio Engineering Medical Institute. 96-well stripped dies were obtained from NUNC Co. The following buffers were used-Covering-100 mmol/l carbonate (pH 9.5), containing 9 g / l NaCl and 0.4 g / l NaN_3 ; Blocking, containing 50 mmol / l Tris-HCl (pH 7.0)), containing 9 g / l NaCl and 0.4 g / l NaN_3 ; and 10 g/l bovine serum albumin, analytical, containing 50 mmol/l TrisHCl (pH 7.8), containing 20 g/l bovine serum albumin, 0.4 g / l NaN_3 , 9 g / l NaCl, and 0.4 ml / l Tween 20, washing buffer 50 mmol / l Tris-HCl (pH 7.4), containing 0.4 g / l NaN_3 , 9 g/l NaCl, and 0.4 ml / l Tween 20. Conjugates of the monoclonal antibody-PAMG-1 and PAMG — 2 with nanogold 30 nm obtained from the company Nano-lab.

Antibodies against HCG and AFP were labeled with Ei^{3+} and Sm^{3+} chelates, respectively.

For this, 1 g / l antibodies in 50 mmol / l carbonate buffer (pH9.5) were mixed with 0.35 mg Ei^{3+} and Sm^{3+} chelate complexes and incubated at 4 °C overnight. Chelate-bound antibodies were purified from unbound reagents on the Sephadex g-25 column (1.5 25 cm) and eluted with a 50 mmol/l Tris-Hcl buffer (pH 7.0) containing 9 g / l —7-

NaCl and 0.5 g / l NaN_3 . The concentration of Ei^{3+} was determined by comparison with the calibrator.

The concentration Ei^{3+} labeled anti — PAMG-1 and PAMG — 2 monoclonal[antibodies calculated by the formula)

$$c = \frac{A_{280} - 0.008c_{Eu^{3+}}}{1.34}$$

Where with A_{280} absorption of Eu^{3+} — labeled anti-PAMG-1 and PAMG — 2 antibodies at 280 Nm, 1.34 is the optical density of 1 g/l EC3-labeled anti-AFP monoclonal antibodies at 280 Nm, and 0.008 is the absorption coefficient of 1 mol/l EC3 chelate at 280 Nm. The yield of the conjugation were obtained by calculating ratio of concentration of Labeled Eu^{3+} monoclonal antibodies of AFP concentration Eu^{3+} in solution. Similarly, the yield of conjugation of Sm^{3+} -labeled antibody to HCG monoclonal antibodies was calculated. One mole of anti-AFP antibodies was labeled with 7.1 mole of Eu^{3+} chelate, while 1 mole of anti-HCG antibodies was labeled with 12.3 moles Of Sm^{3+} chelate. Standard concentrations of Eu^{3+} and Sm^{3+} , (5 g / L) and optimal key settings of ICP-MS equipment were selected for optimal analysis. The dependence of the signal intensities Eu^{3+} and Sm^{3+} , were investigated in the argon flow at flow rates of 0.7-1.1 l / min. Eu^{3+} and Sm^{3+} , behaved similarly giving a total peak at 1 l / min (Fig. 1). The maximum intensity of Eu^{3+} was twice as high as Sm^{3+} , (52.2% for Eu^{3+} and 26.7% for Sm^{3+}).

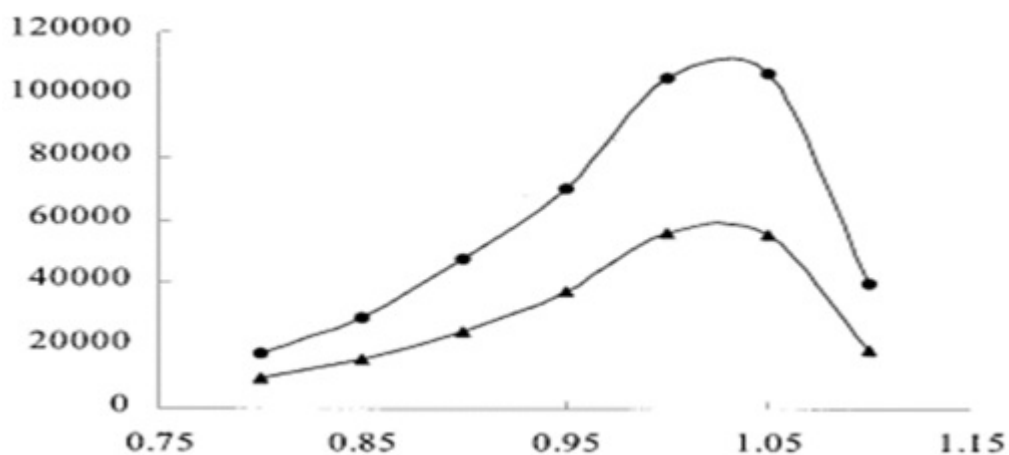


Fig 1. Optimization of argon flow to analyze IPMS-ELISA On the x-axis shows the flow of argon(l / min) On the ordinate axis-the power radiation of the isotope. Concentration of Eu^{3+} and Sm^{3+} , 5g/ l, current power 1250 W, voltage 8 V

Two definitions were performed for each sample. Analytical results are obtained by calculating the average of two definitions of each sample. The ratio is calculated by the linear least squares method according to the criterion

Student. The calibration curves for IPMS-IFA are shown in Fig.2

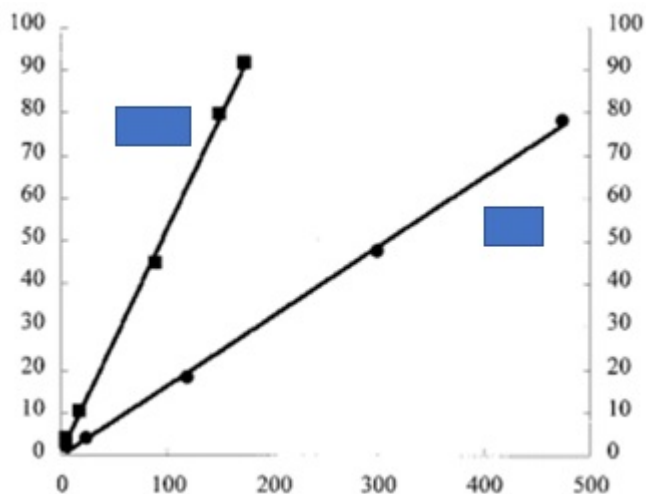


Fig.2 Calibration curve for concentration determination

conjugates along the abscissa axis of concentration of PAMG-1 and PAMG-2 in ng/ml.

On the ordinate axis the ratio of signals is $(^{52}\text{Sm}/^{174}\text{Yb})\%$

Blood samples were taken from 20 patients of a network of clinics "Family doctor" (Moscow, Russian Federation.) after obtaining informed consent from patients. Serum was obtained as previously described [3] serum Samples were then stored at 20 °C under nitrogen gas prior to use

IPMS-ELISA was performed according to the standard procedure for ELISA [3], the complex MAT PAMG-1 and Ei^{3+} and PAMG — 2 with Sm^{3+} were used as conjugates . After washing from unbound conjugates, the label was washed with a solution of HNO_3 (10 ml/l) and delivered to the mass spectrometer by a peristaltic pump. The analysis was performed after optimization of the reaction

Calibration Curve PAMG-1 and PAMG-2 Vaya and sensitivity analysis are shown in Fig.2 simultaneous determination of PAMG-1 and was in the concentration range 4.6-500 ng / ml, and for PAMG-2 ng / ml. regression Equations for calibration curves were: $I=0.16 C+0.06$ for AFP($r=0.9984$) and

$I=0.52 C+0.96$ for PAMG — 1 ($R^2=0.9987$), where I is the relative signal intensity and C is the concentration. The lower limit was determined as 3SD lower than the mean of 12 measurements and was 1.2 $\mu\text{g/l}$ for PAMG-1 1.7 $\mu\text{g/l}$ for PAMG-2 IHA for the two antigens was performed as previously described [3] as kongugatov used conjugates of the MAT against AFP and beta HCG, labeled with nanogold particle size of 30nm. The results were taken into account on the hardware-software complex "Expert-lab".

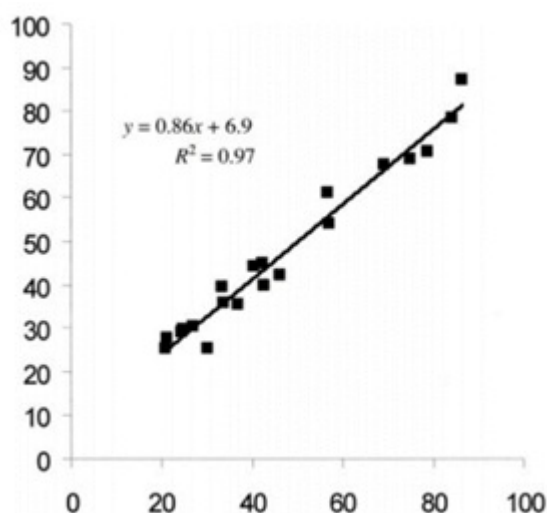


Fig3 the relationship between the results of determining the concentrations of the two encountered

in the sera of patients containing PAMG-1 and PAMG — 2 methods ELISA and IPMS-ELISA for PAMG-1 and PAMG-2. On the ordinate axis, AND on the abscissa axis, the results of IPMS-ELISA Showed that the relationship between the results of determining the concentrations of two oncoantigens in the serum of patients containing PAMG-1 and PAMG-2 the methods of ICA and IPMS-ELISA is very high and practically insignificant, which of these methods is used for diagnosis. The advantage of IPMS-ELISA is its high potential for automation, faster than traditional ELISA execution at a relative cheapness[4,5]. The disadvantage of the method is the work with radioactive isotopes, which is often organizationally difficult due to environmental insecurity. On the other hand, IHA is faster, not more expensive, but less amenable to automation. Thus, for the screening of large arrays of samples, IPMS-IFA is preferable, and for home screening of IHA .

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